

New Hampshire Volunteer Lake Assessment Program

2002 Bi-Annual Report for Cobbetts Pond Windham



NHDES
Water Division
Watershed Management Bureau
6 Hazen Drive
Concord, NH 03301



OBSERVATIONS & RECOMMENDATIONS

After reviewing data collected from **COBBETTS POND**, the program coordinators recommend the following actions.

We would like to encourage your monitoring group to conduct more sampling events in the future. Typically we recommend that each monitoring group sample at least three times per summer (once in June, July, and August). We understand that the number of sampling events you decide to conduct per summer will depend upon volunteer availability and your associations' water monitoring goals and funding availability. However, with a limited amount of data it is difficult to determine accurate and representative lake quality trends. Since weather patterns and activity in the watershed can change throughout the summer, and from year to year (and even from hour to hour during a rain event), it is a good idea to sample more than once or twice over the course of the season. If you are having difficulty finding volunteers to help sample, or to pick-up or drop-off equipment at one of the labs, please give the VLAP Coordinator a call and we will try to help you work out an arrangement.

FIGURE INTERPRETATION

- **Figure 1 and Table 1:** The graphs in Figure 1 (Appendix A) show the historical and current year chlorophyll-a concentration in the water column. Table 1 (Appendix B) lists the maximum, minimum, and mean concentration for each sampling season that the lake/pond has been monitored through the program.

Chlorophyll-a, a pigment naturally found in plants, is an indicator of the algal abundance. Because algae are usually microscopic plants that contain chlorophyll-a, and are naturally found in lake ecosystems, the chlorophyll-a concentration found in the water gives an estimation of the concentration of algae or lake productivity. The mean (average) summer chlorophyll-a concentration for New Hampshire's lakes and ponds is 7.02 ug/L.

Similar to the summer of 2001, the summer of 2002 was filled with many warm and sunny days and there was a lower than normal

amount of rainfall during the latter-half of the summer. The combination of these factors resulted in relatively warm surface waters throughout the state. The lack of fresh water to the lakes/ponds reduced the rate of flushing which may have resulted in water stagnation. Due to these conditions, many lakes and ponds experienced increased algae growth, including filamentous green algae (the billowy clouds of green algae typically seen floating near shore), and some lakes/ponds experienced nuisance cyanobacteria (blue-green algae) blooms.

STATION 1:

The current year data (the top graph) show that the chlorophyll-a concentration **increased slightly** from July to September.

The historical data (the bottom graph) show that the 2002 chlorophyll-a mean is **less than** the state mean.

Overall, the statistical analysis of the historical data show that the chlorophyll-a concentration has **significantly increased** since monitoring began in **1988**. Specifically, the chlorophyll-a concentration has **increased** (meaning **worsened**) on average by **approximately 4.5 percent per** sampling season during the sampling period **1988 to 2002**. (Note: Please refer to Appendix E for the statistical analysis explanation and data print out.)

STATION 2:

The current year data (the top graph) show that the chlorophyll-a concentration **decreased slightly** from July to September.

The historical data (the bottom graph) show that the 2002 chlorophyll-a mean is **less than** the state mean.

Overall, the statistical analysis of the historical data (the bottom graph) shows that the mean annual chlorophyll-a concentration has **not significantly changed** (either *increased* or *decreased*) since monitoring began in **1988**. Specifically, the chlorophyll-a concentration has **varied** (meaning **fluctuated**), but has not *continually increased* or *decreased* during the sampling period 1988 – 2002. (Note: Please refer to Appendix E for the detailed statistical analysis explanation and data print out.)

While algae are naturally present in all lakes/ponds, an excessive or increasing amount of any type is not welcomed. In freshwater lakes/ponds, phosphorus is the nutrient that algae depend upon for growth. Therefore, algal concentrations may increase when there is an increase in nonpoint sources of nutrient loading from the watershed, or in-lake sources of phosphorus loading (such as phosphorus releases from the sediments). It is important to

continually educate residents about how activities within the watershed can affect phosphorus loading and lake quality.

- **Figure 2 and Table 3:** The graphs in Figure 2 (Appendix A) show historical and current year data for lake/pond transparency. Table 3 lists the maximum, minimum and mean transparency data for each sampling season that the lake/pond has been monitored through the program.

Volunteer monitors use the Secchi-disk, a 20 cm disk with alternating black and white quadrants, to measure water clarity (how far a person can see into the water). Transparency, a measure of water clarity, can be affected by the amount of algae and sediment from erosion, as well as the natural colors of the water. The mean (average) summer transparency for New Hampshire's lakes and ponds is 3.7 meters.

Two different weather related patterns occurred this past spring and summer that influenced lake quality during the summer season.

In late May and early June of 2002, numerous rainstorms occurred. Stormwater runoff associated with these rainstorms may have increased phosphorus loading, and the amount of soil particles washed into waterbodies throughout the state. Some lakes and ponds experienced lower than typical transparency readings during late May and early June.

However, similar to the 2001 sampling season, the lower than average amount of rainfall and the warmer temperatures during the latter-half of the summer resulted in a few lakes/ponds reporting their best-ever Secchi-disk readings in July and August (a time when we often observe reduced clarity due to increased algal growth)!

STATION 1:

The current year data (the top graph) show that the in-lake transparency **increased very slightly** from July to September.

The historical data (the bottom graph) show that the 2002 mean transparency is **slightly greater than** the state mean.

Overall, the statistical analysis of the historical data show that the transparency has **significantly decreased** since monitoring began in **1988**. Specifically, the in-lake transparency has **decreased (meaning worsened)** on average by **approximately 2.5 percent** per sampling season during the sampling period **1988 to 2002**. (Note: Please refer to Appendix E for the statistical analysis explanation and data print out.)

STATION 2:

The current year data (the top graph) show that the in-lake transparency **increased slightly** from July to September.

The historical data (the bottom graph) show that the 2002 mean transparency is **slightly less than** the state mean.

Overall, the statistical analysis of the historical data show that the transparency has **significantly decreased** since monitoring began in **1988**. Specifically, the in-lake transparency has **decreased (meaning worsened)** on average by **approximately 2.0 percent** per sampling season during the sampling period **1988** to **2002**. (Note: Please refer to Appendix E for the statistical analysis explanation and data print out.)

Typically, high intensity rainfall causes erosion of sediments into lakes/ponds and streams, thus decreasing clarity. Efforts should continually be made to stabilize stream banks, lake/pond shorelines, disturbed soils within the watershed, and especially dirt roads located immediately adjacent to the edge of tributaries and the lake/pond. Guides to Best Management Practices designed to reduce, and possibly even eliminate, nonpoint source pollutants, such as sediment loading, are available from NHDES upon request.

- **Figure 3 and Table 8:** The graphs in Figure 3 (Appendix A) show the amounts of phosphorus in the epilimnion (the upper layer) and the hypolimnion (the lower layer); the inset graphs show current year data. Table 8 (Appendix B) lists the annual maximum, minimum, and median concentration for each deep spot layer and each tributary since the lake/pond has joined the program.

Phosphorus is the limiting nutrient for plant and algae growth in New Hampshire's freshwater lakes and ponds. Too much phosphorus in a lake/pond can lead to increases in plant and algal growth over time. The median summer total phosphorus concentration in the epilimnion (upper layer) of New Hampshire's lakes and ponds is 11 ug/L. The median summer phosphorus concentration in the hypolimnion (lower layer) is 14 ug/L.

STATION 1:

The current year data for the epilimnion (the top inset graph) show that the total phosphorus concentration **decreased** from July to September.

The historical data show that the 2002 mean epilimnetic total phosphorus concentration is **approximately equal to** the state mean.

The current year data for the hypolimnion (the bottom inset graph) show that the total phosphorus concentration in September was much **greater than** the state median.

It is important to note that the July hypolimnetic phosphorus result was removed from that data set. The total phosphorus concentration in July hypolimnion sample was **very high** compared to the historic data (134 ug/L), and the turbidity of the sample was also **very high** (15.7 NTUs). This suggests that the lake/pond bottom may have been disturbed by the anchor or by the Kemmerer Bottle while sampling. When the lake/pond bottom is disturbed, sediment, which typically contains attached phosphorus, is released into the water column. When collecting the hypolimnion sample, please check to make sure that there is no sediment in the Kemmerer Bottle before filling the sample bottles.

Overall, the statistical analysis of the historical data show that the total phosphorus concentration in the epilimnion (upper layer) has **not significantly changed** (either *increased* or *decreased*) since monitoring began in **1988**. Specifically, the total phosphorus concentration in the epilimnion has remained **relatively stable**. (Note: Please refer to Appendix E for the statistical analysis explanation and data print out.)

Overall, the statistical analysis of the historical data show that the phosphorus concentration in the hypolimnion (lower layer) has **significantly increased** since monitoring began in **1988**. Specifically, the phosphorus concentration in the hypolimnion has increased on average at a rate of approximately **7.3 percent** per season during the sampling period **1988** to **2002** (Please refer to Appendix F for the statistical analysis explanation and data print out).

STATION 2:

The current year data for the epilimnion (the top inset graph) show that the total phosphorus concentration **increased slightly** from July to September.

The historical data show that the 2002 mean epilimnetic total phosphorus concentration is **slightly less than** the state mean.

The current year data for the hypolimnion (the bottom inset graph) show that the total phosphorus concentration **increased greatly** from July to September.

The historical data show that the 2002 mean hypolimnetic total phosphorus concentration is **much greater than** the state mean.

It is important to note that the turbidity of the hypolimnion sample was **very high** (26.8 NTUs). This suggests that the lake/pond bottom may have been disturbed by the anchor or by the Kemmerer Bottle while sampling. However, this data point was not removed from the data set because the total phosphorus concentration was not significantly greater than the historic total phosphorus data for the hypolimnion.

Overall, the statistical analysis of the historical data show that the total phosphorus concentration in the epilimnion (upper layer) has **not significantly changed** (either *increased* or *decreased*) since monitoring began in **1988**. Specifically, the total phosphorus concentration in the epilimnion has remained **relatively stable**. (Note: Please refer to Appendix E for the statistical analysis explanation and data print out.)

Overall, the statistical analysis of the historical data show that the phosphorus concentration in the hypolimnion (lower layer) has **significantly increased** since monitoring began in **1988**. Specifically, the phosphorus concentration in the hypolimnion has increased on average at a rate of approximately **7.3 percent** per season during the sampling period **1988 to 2002** (Please refer to Appendix F for the statistical analysis explanation and data print out).

One of the most important approaches to reducing phosphorus loading to a waterbody is to continually educate watershed residents about its sources and how excessive amounts can adversely impact the ecology and value of lakes and ponds. Phosphorus sources within a lake or pond's watershed typically include septic systems, animal waste, lawn fertilizer, road and construction erosion, and natural wetlands. If you would like to educate watershed residents about how they can help to reduce phosphorus loading into the lake/pond, please contact the VLAP Coordinator.

TABLE INTERPRETATION

➤ Table 2: Phytoplankton

Table 2 lists the current and historic phytoplankton species observed in the lake/pond.

The dominant phytoplankton species observed this year at **STATION 1** were ***Fragilaria* (a diatom), *Asterionella* (a diatom), and *Dinobryon* (a golden-brown algae).**

The dominant phytoplankton species observed this year at **STATION 2** were ***Fragilaria* (a diatom), *Dinobryon* (a golden-brown algae), and *Tabellaria* (a diatom).**

Phytoplankton populations undergo a natural succession during the growing season (Please refer to page 12 of the “Biological Monitoring Parameters” section of this report for a more detailed explanation regarding seasonal plankton succession). Diatoms and golden-brown algae are typical in New Hampshire’s less productive lakes and ponds. An overabundance of cyanobacteria (previously referred to as blue-green algae) indicates that there may be an excessive total phosphorus concentration in the lake/pond, or that the ecology is out of balance.

➤ **Table 2: Cyanobacteria (Blue-green algae)**

Small amounts of the cyanobacterium *Anabaena* was observed in the plankton sample at both deep spots this season. ***This species, if present in large amounts, can be toxic to livestock, wildlife, pets, and humans.*** Cyanobacteria can reach nuisance levels when excessive nutrients and favorable environmental conditions occur. As with the summer of 2001, we observed that some lakes and ponds had cyanobacteria present during the 2002 summer season, likely due to the many warm and sunny days that occurred this summer, which may have accelerated algal and bacterial growth. In addition, the lower than normal amount of rainfall during the latter half of the summer, meant that the slow flushing rates resulted in less phosphorus exiting the lake outlet and more phosphorus being available for plankton growth.

The presence of cyanobacteria serves as a reminder of the lake’s/pond’s delicate balance. Watershed residents should continue to act proactively to reduce nutrient loading into the lake/pond by eliminating fertilizer use on lawns, keeping the lake/pond shoreline natural, re-vegetating cleared areas within the watershed, and properly maintaining septic systems and roads.

In addition, residents should also observe the lake/pond in September and October during the time of fall turnover (lake mixing) to document any algal blooms that may occur. Cyanobacteria (blue-green algae) have the ability to regulate their depth in the water column by producing or releasing gas from vesicles. However, occasionally lake mixing can affect their buoyancy and cause them to rise to the surface and bloom. Wind and currents tend to “pile” cyanobacteria into scums that accumulate in one section of the lake/pond. If a fall bloom occurs, please contact the VLAP Coordinator.

➤ **Table 4: pH**

Table 4 (Appendix B) presents the in-lake and tributary current year and historical pH data.

pH is measured on a logarithmic scale of 0 (acidic) to 14 (basic). pH is important to the survival and reproduction of fish and other aquatic life. A pH below 5.5 severely limits the growth and reproduction of fish. A pH between 6.5 and 7.0 is ideal for fish. The mean pH value for the epilimnion (upper layer) in New Hampshire's lakes and ponds is 6.5, which indicates that the surface waters in state are slightly acidic. For a more detailed explanation regarding pH, please refer to page 16 of the "Chemical Monitoring Parameters" section of this report.

The mean pH at the **STATION 1** deep spot this season ranged from **6.64** in the hypolimnion to **7.21** in the epilimnion. The mean pH at the **STATION 2** deep spot this season ranged from **6.70** in the hypolimnion to **7.32** in the epilimnion. This means that the pH of the water column ranged from being *slightly acidic* near the bottom of the lake to *slightly basic (meaning alkaline)* near to surface of the lake.

Due to the presence of granite bedrock in the state and the deposition of acid rain, there is not much that can be done to effectively increase lake/pond pH.

➤ **Table 5: Acid Neutralizing Capacity**

Table 5 in Appendix B presents the current year and historic epilimnetic ANC for each year the lake/pond has been monitored through VLAP.

Buffering capacity or ANC describes the ability of a solution to resist changes in pH by neutralizing the acidic input to the lake. For a more detailed explanation, please refer to page 16 of the "Chemical Monitoring Parameters" section of this report.

The Acid Neutralizing Capacity (ANC) of the epilimnion (the upper layer) at both deep spot stations continues to be *very high (approximately 21 to 23 mg/L as CaCO₃, and much greater than* the state mean of 6.7 mg/L (Table 5). Specifically, this means that the lake/pond is *"not vulnerable"* to acidic inputs (such as acid precipitation).

➤ **Table 6: Conductivity**

Table 6 in Appendix B presents the current and historic conductivity values for tributaries and in-lake data. Conductivity is the numerical expression of the ability of water to carry an electric current. For a more detailed explanation, please refer to page 16 of the "Chemical Monitoring Parameters" section of this report.

The conductivity has **increased** in the lake/pond and inlets since monitoring began (Table 6). Typically, sources of increased conductivity are due to human activity. These activities include septic systems that fail and leak leachate into the groundwater (and eventually into the tributaries and the lake/pond), agricultural runoff, and road runoff (which contains road salt during the spring snow melt). New development in the watershed can alter runoff patterns and expose new soil and bedrock areas, which could contribute to increasing conductivity. In addition, natural sources, such as iron deposits in bedrock, can influence conductivity. It is likely that the high conductivity level in the lake is due to the proximity of the interstate and the use of salt on the roadways in the winter.

We recommend that your monitoring group conduct stream surveys and stormwater sampling along the inlets with elevated conductivity so that we can more accurately “pin-point” what may be causing the increases. For a detailed explanation on how to conduct a stream survey and stormwater sampling, please refer to this year’s “Special Topic Article” which is included in Appendix D of this report.

➤ **Table 8: Total Phosphorus**

Table 8 in Appendix B presents the current year and historic total phosphorus data for in-lake and tributary stations. Phosphorus is the nutrient that limits the algae’s ability to grow and reproduce. Please refer to page 17 of the “Chemical Monitoring Parameters” section of this report for a more detailed explanation.

➤ **Table 9 and Table 10: Dissolved Oxygen and Temperature Data**

Table 9 in Appendix B shows the dissolved oxygen/temperature profile(s) for the 2002 sampling season. Table 10 shows the historical and current year dissolved oxygen concentration in the hypolimnion (lower layer). The presence of dissolved oxygen is vital to fish and amphibians in the water column and also to bottom-dwelling organisms. Please refer to the “Chemical Monitoring Parameters” section of this report for a more detailed explanation.

The dissolved oxygen concentration was **low in the hypolimnion** at the deep spot of the lake/pond. As stratified lakes/ponds age, oxygen becomes **depleted** in the hypolimnion (the lower layer) by the process of decomposition. Specifically, the loss of oxygen in the hypolimnion results primarily from the process of biological breakdown of organic matter (i.e.; biological organisms use oxygen to break down organic matter), both in the water column and particularly at the bottom of the lake/pond where the water meets the sediment.

In addition, during this season, and many past sampling seasons the lake/pond has had a lower dissolved oxygen concentration and a

higher total phosphorus concentration in the hypolimnion (the lower layer) than in the epilimnion (the upper layer). These data suggest that the process of **internal total phosphorus loading** (commonly referred to as **internal loading**) is occurring in the lake/pond. When oxygen levels are depleted to less than 1 mg/L in the hypolimnion, the phosphorus that is normally bound up with metals in the sediment may be re-released into the water column. Depleted oxygen concentration in the hypolimnion of thermally stratified lakes/ponds typically occurs as the summer progresses.

The **low** oxygen level in the hypolimnion is a sign of the lake's/pond's **aging** and **declining** health. This year the DES biologist conducted the temperature/dissolved oxygen profile in **September**. We recommend that the annual biologist visit for the 2003 sampling season be scheduled during **June** so that we can determine if oxygen is depleted in the hypolimnion **earlier** in the sampling season.

➤ **Table 11: Turbidity**

Table 11 in Appendix B lists the current year and historic data for in-lake and tributary turbidity. Turbidity in the water is caused by suspended matter, such as clay, silt, and algae. Water clarity is strongly influenced by turbidity. Please refer to page 19 of the "Other Monitoring Parameters" section of this report for a more detailed explanation.

As discussed previously, the turbidity of the hypolimnion (lower layer) sample was elevated on a couple of occasions this season. This suggests that the lake/pond bottom may have been disturbed by the anchor or by the Kemmerer Bottle while sampling. When the lake/pond bottom is disturbed, sediment, which typically contains attached phosphorus, is released into the water column. When collecting the hypolimnion sample, please check to make sure that there is no sediment in the Kemmerer Bottle before filling the sample bottles.

➤ **Table 12: Bacteria (*E.coli*)**

Table 12 lists the current year data for bacteria (*E.coli*) testing. *E. coli* is a normal bacterium found in the large intestines in humans and other warm-blooded animals. *E.coli* is used as an indicator organism because it is easily cultured, and its presence in the water, in defined amounts, indicates that sewage **MAY** be present. If sewage is present in the water, potentially harmful pathogens may also be present. Please consult page 20 of the "Other Monitoring Parameters" section of the report for the current standards for *E. coli* in surface waters. If residents are concerned about sources of *E.coli* such as septic system impacts, animal waste, or waterfowl waste, it is best to conduct *E. coli* testing when the water table is high or after rain events.

The *E. coli* result was very low on each sampling event at the **Town Beach** this season. Specifically, results were **7** counts or less, which is ***well below*** the state standard of 88 counts per 100 mL for designated public beaches. If you are concerned about *E. coli* levels at this beach, you may want to repeat this test on a weekend during heavy beach use or after a rain event. Since bacteria die quickly in cool pond waters, testing is most accurate and most representative of the health risk to bathers when the source (humans, animals, or waterfowl) is present.

The *E. coli* concentration at **Castleton Brook** was elevated in **July** (200 counts per 100 mL). However, the concentration ***was not above*** the state standard of 406 counts per 100 mL designated for Class B waters. If you are concerned about *E. coli* levels at this station, your monitoring group may want to conduct stormwater sampling in this area so that we can determine what may be causing the increases. For a detailed explanation on how to conduct stormwater sampling, please refer to this year's special topic which is found in Appendix D of this report.

DATA QUALITY ASSURANCE AND CONTROL

Annual Assessment Audit:

During the annual visit to your lake/pond, the biologist conducted a "Sampling Procedures Assessment Audit" for your monitoring group. Specifically, the biologist observed the performance of your monitoring group while sampling and filled out an assessment audit sheet to document the ability of the volunteer monitors to follow the proper field sampling procedures (as outlined in the VLAP Monitor's Field Manual). This assessment is used to identify any aspects of sample collection in which volunteer monitors are not following the proper procedures, and also provides an opportunity for the biologist to retrain the volunteer monitors as necessary. This will ultimately ensure that the samples that the volunteer monitors collect are truly representative of actual lake and tributary conditions.

Overall, your monitoring group performed ***very well*** while collecting samples on the annual biologist visit this season! Specifically, the members of your monitoring group followed the majority of the proper field sampling procedures. The biologist did identify a few aspects regarding sample collection that the volunteer monitors could improve upon. They are as follows:

- **Finding the deep spot:** Please remember to locate the deep spot using three reference points from the shoreline. This method is

known as **triangulation**. In addition, depth finders and Global Positioning System (GPS) technology may be used to further pinpoint the location of the deep spot. In addition, please remember to check the depth of the deep spot by **sounding** to ensure that you have actually located the deepest spot. To sound the bottom, simply fill the Kemmerer bottle with lake water from the surface and then lower the bottle into the lake until you feel it touch the bottom. When you have reached the bottom, check the depth on the calibrated chain. You may need to move to another location and repeat this procedure a few times until the deepest spot is located. When you have found the deep spot, please remember to write the depth of the field data sheet. Please note, sounding may disturb the sediment, so please allow the bottom to settle out before collecting the deepest sample.

- **Anchoring at deep spot:** Please remember to use an anchor with sufficient weight and sufficient amount of rope to prevent the boat from drifting while sampling at the deep spot. It is difficult for the biologist to collect an accurate and representative dissolved oxygen/temperature profile when the boat is drifting. In addition, it is difficult to view the secchi disk and collect samples from the proper depths when the boat is drifting. Depending on the depth of the lake/pond and the wind conditions, it may be necessary to use two anchors!

Sample Receipt Checklist:

Each time your monitoring group dropped off samples at the laboratory this summer, the laboratory staff completed a sample receipt checklist to assess and document if the volunteer monitors followed proper sampling techniques when collecting the samples. The purpose of the sample receipt checklist is to minimize, and hopefully eliminate, future re-occurrences of improper sampling techniques.

Overall, the sample receipt checklist showed that your monitoring group did a **very good** job when collecting samples this season! Specifically, the members of your monitoring group followed the majority of the proper field sampling procedures when collecting and submitting samples to the laboratory. However, the laboratory did identify one aspect of sample collection that the volunteer monitors could improve upon.

- **Sample Labels:** On the **July 15th** sampling event, at least one sample bottle was not labeled. However, by process of elimination or by contacting the volunteer monitors, it was possible for the lab to determine which sample bottles corresponded to what sampling locations. Please make sure to label your samples with a waterproof pen (a black sharpie permanent marker works best), preferably before

sampling. If your association has made its own sample bottle labels, please make sure to fold over one corner of each label before placing it on a sample bottle so that the label will not become permanently attached to the bottle. In addition, please make sure that the labels will stick to the bottles when they are wet.

OTHER COMMENTS

- The tributaries to the pond were not sampled for conductivity, pH, phosphorus, or turbidity this season. (The inlets were only sampled for *E.coli*). We recommend that your monitoring group sample all of the inlets during the late spring, soon after the snow has melted, for the standard parameters so that we can determine the quality of the water that flows into the pond.
- Since the interstate may be widened in this area in the near future, it would be a good idea to conduct stream surveys and stormwater sampling for all of the inlets to the pond. This type of data and information may become useful if roadway expansion construction activities appear to be causing water quality problems in the pond or inlets. For a detailed explanation on how to conduct a stream survey and stormwater sampling, please refer to this year's "Special Topic Article" which is included in Appendix D of this report.

NOTES

- **Biologist's Note (9/6/02):** A lot of drifting occurred while at the deep spot. Therefore, the dissolved oxygen profile is not truly representative at some depths. Please use an anchor while at the deep spot. Very obvious sulfur smell in hypolimnion sample at both stations.

USEFUL RESOURCES

Changes to the Comprehensive Shoreland Protection Act: 2001 Legislative Session, NHDES Fact Sheet, (603) 271-3505, or www.des.state.nh.us/factsheets/sp/sp-8.htm

Cyanobacteria in New Hampshire Waters Potential Dangers of Blue-Green Algae Blooms, NHDES Fact Sheet, (603) 271-3505, or www.des.state.nh.us/factsheets/wmb/wmb-10.htm

The Lake Pocket Book. Prepared by The Terrene Institute, 2000. (internet: www.terrene.org, phone 800-726-4853)

Managing Lakes and Reservoirs, Third Edition, 2001. Prepared by the North American Lake Management Society (NALMS) and the Terrene Institute in cooperation with the U.S. Environmental Protection Agency. Copies are available from NALMS (internet: www.nalms.org, phone 608-233-2836), and the Terrene Institute (internet: www.terrene.org, phone 800-726-4853)

Organizing Lake Users: A Practical Guide. Written by Gretchen Flock, Judith Taggart, and Harvey Olem. Copies are available from the Terrene Institute (internet: www.terrene.org, phone 800-726-4853)

Proper Lawn Care in the Protected Shoreland: The Comprehensive Shoreland Protection Act, WD-SP-2, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/sp/sp-2.htm

Sand Dumping - Beach Construction, WD-BB-15, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/bb/bb-15.htm

Swimmers Itch, WD-BB-2, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/bb/bb-2.htm

Use of Lakes or Streams for Domestic Water Supply, WD-WSEB-1-11, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/ws/ws-1-11.htm

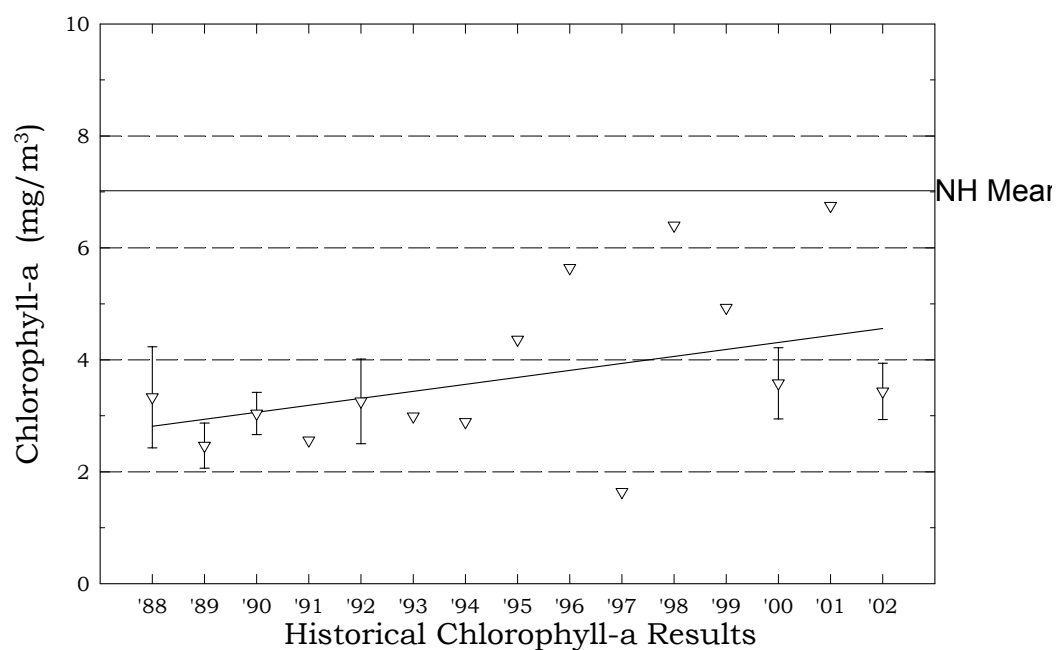
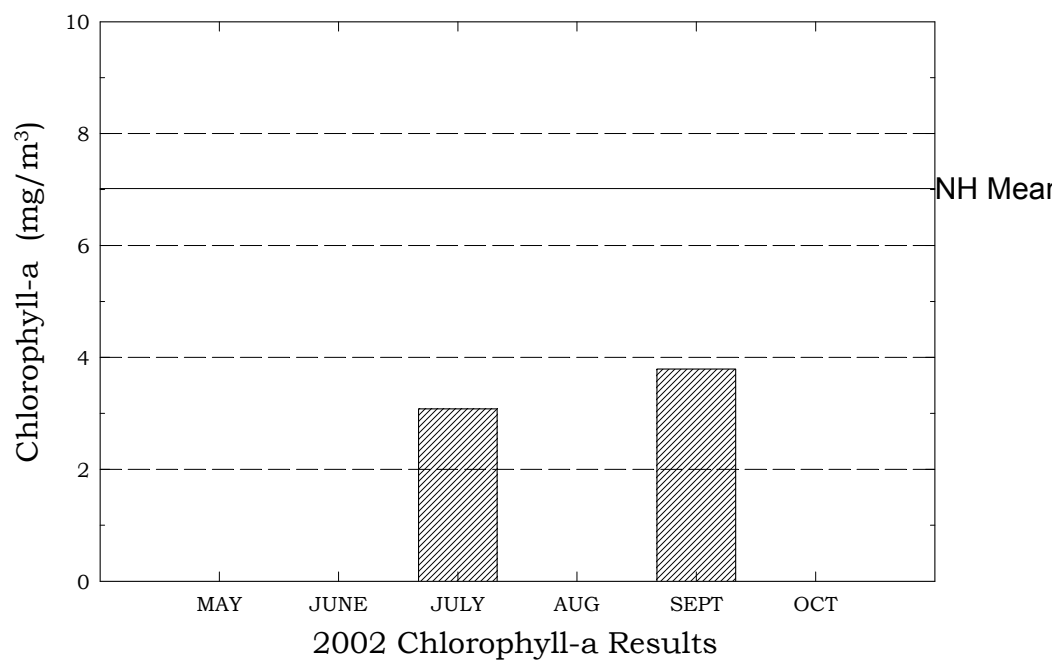
Water Milfoil, WD-BB-1, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/bb/bb-1.htm

Weed Watchers: An Association to Halt the Spread of Exotic Aquatic Plants, WD-BB-4, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/bb/bb-4.htm

Appendix A: Graphs

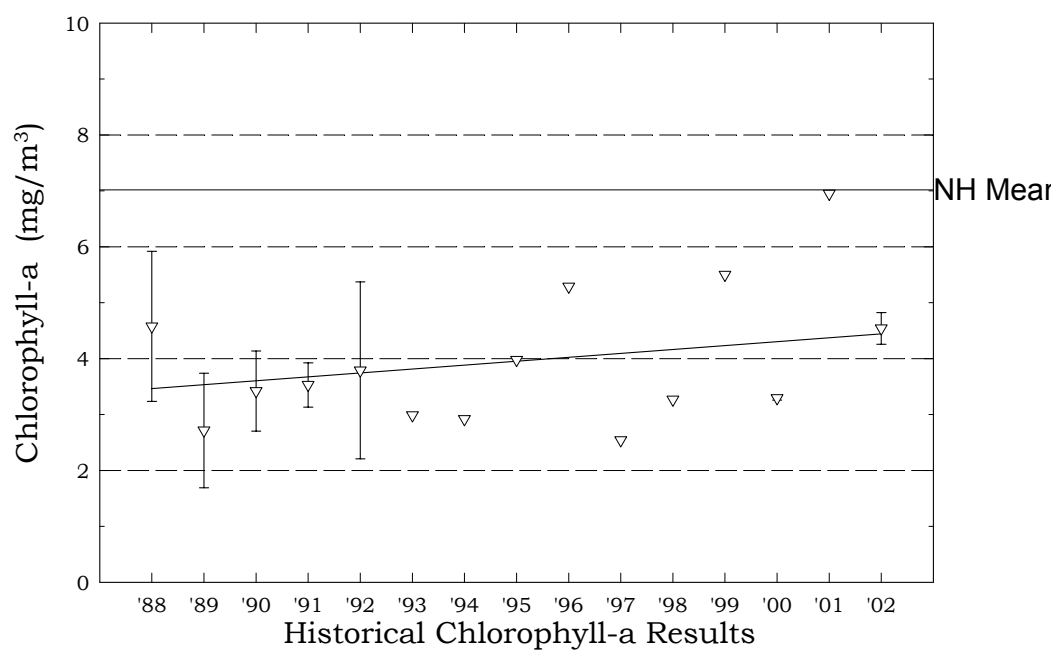
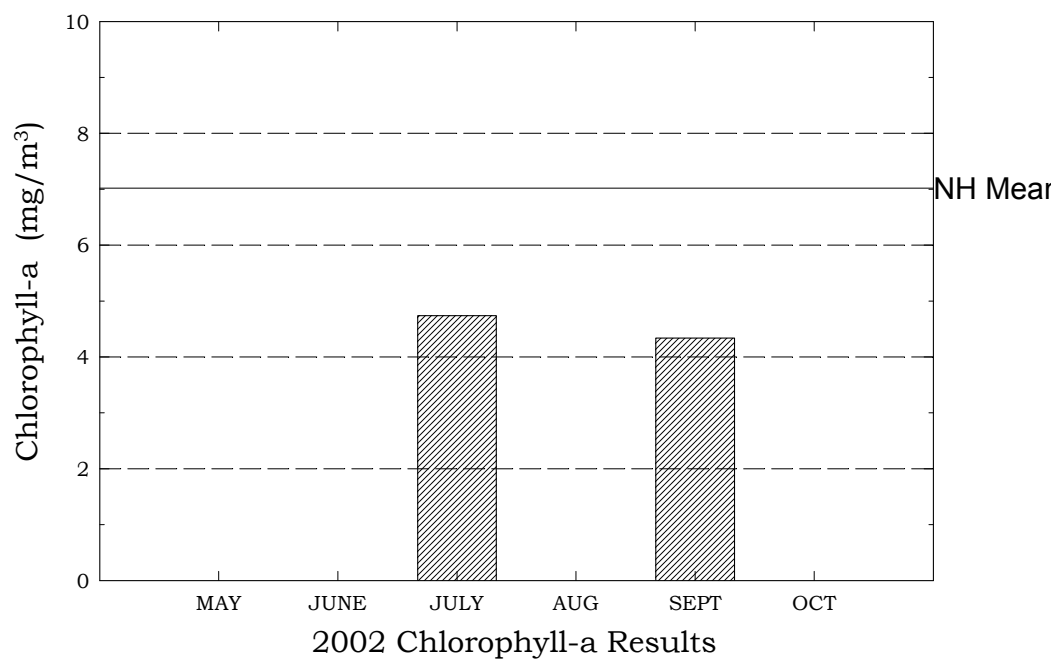
Cobbetts Pond, Windham Station 1

Figure 1. Monthly and Historical Chlorophyll-a Results



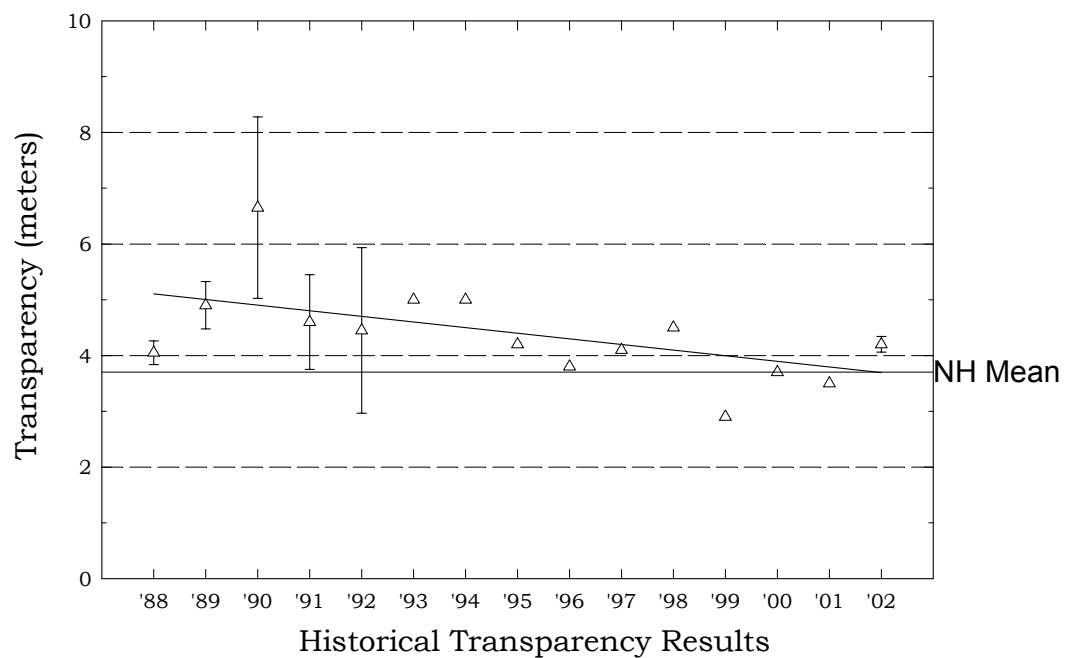
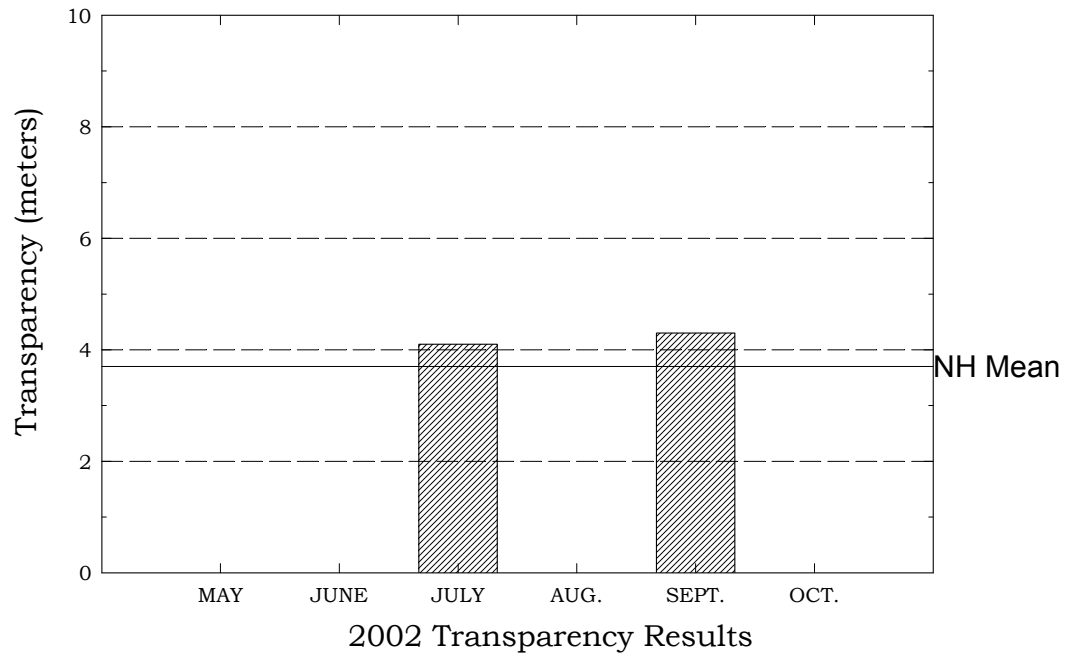
Cobbetts Pond, Windham Station 2

Figure 1. Monthly and Historical Chlorophyll-a Results



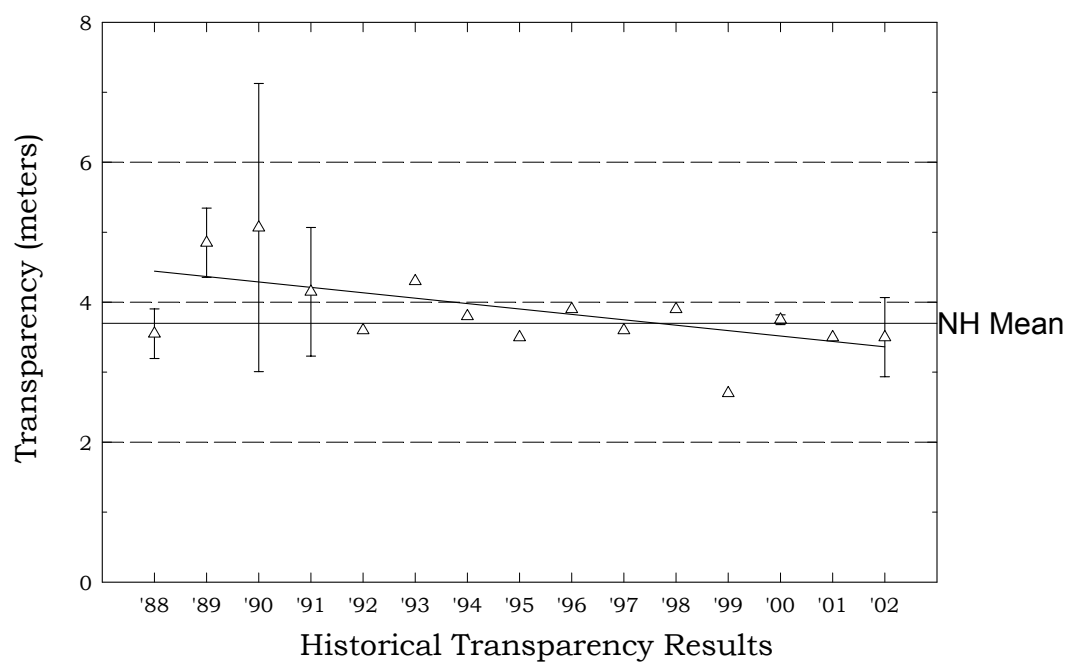
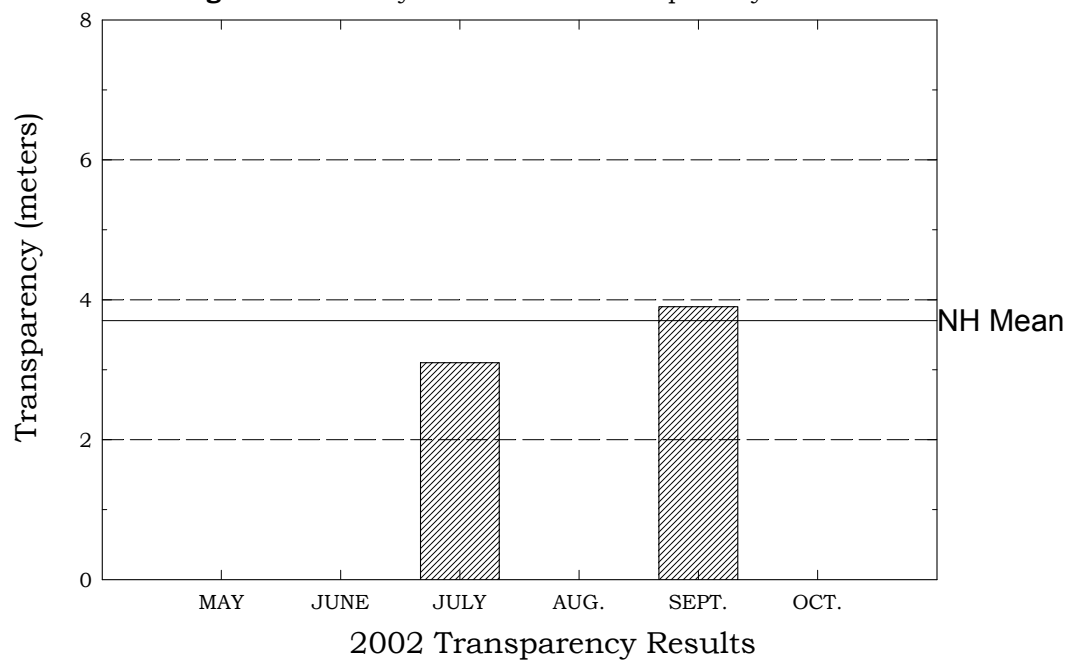
Cobbetts Pond, Windham Station 1

Figure 2. Monthly and Historical Transparency Results



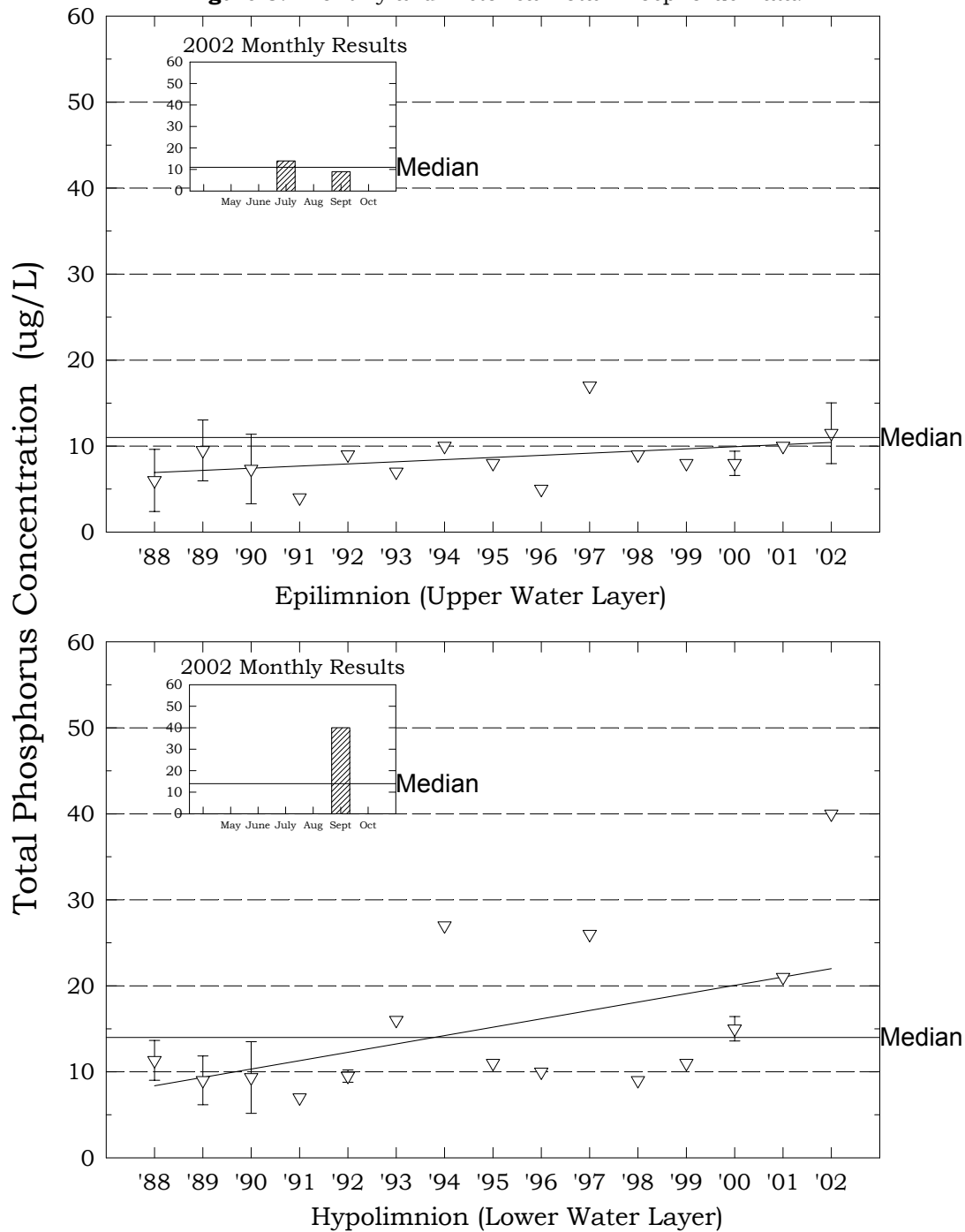
Cobbetts Pond, Windham Station 2

Figure 2. Monthly and Historical Transparency Results



Cobbetts Pond, Station 1, Windham

Figure 3. Monthly and Historical Total Phosphorus Data.



Cobbetts Pond, Station 2, Windham

Figure 3. Monthly and Historical Total Phosphorus Data.

